

**DESIGN AND SYNTHESIS OF 2-[4-[4-(*m*-(ETHYLSULFONAMIDO)-  
PHENYL)PIPERAZIN-1-YL]BUTYL]-1,3-DIOXOPERHYDROPYRROLO[1,2-  
*c*]IMIDAZOLE (EF-7412) USING NEURAL NETWORKS. A SELECTIVE  
DERIVATIVE WITH MIXED 5-HT<sub>1A</sub>/D<sub>2</sub> ANTAGONIST PROPERTIES**

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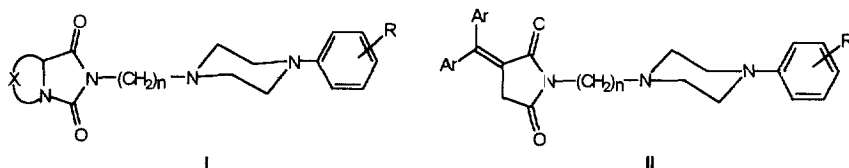
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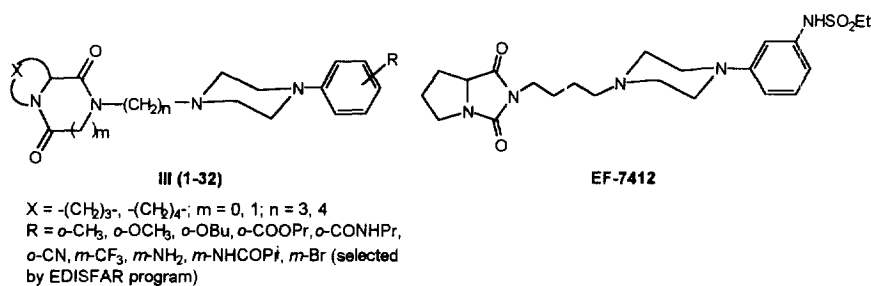
**Abstract:** A test series of 32 phenylpiperazines **III** with affinity for 5-HT<sub>1A</sub> and  $\alpha_1$  receptors was subjected to QSAR analysis using artificial neural networks (ANNs), in order to get insight into the structural requirements that are responsible for 5-HT<sub>1A</sub>/ $\alpha_1$  selectivity. Good models and predictive power were obtained for 5-HT<sub>1A</sub> and  $\alpha_1$  receptors. A comparison of these models gives information for the design of the new ligand **EF-7412** (5-HT<sub>1A</sub>:  $K_i$  (nM) = 27;  $\alpha_1$ :  $K_i$  (nM) > 1000). This derivative displayed affinity for dopamine D<sub>2</sub> receptor ( $K_i$  = 22 nM) and is selective for all other receptor examined (5-HT<sub>2A</sub>, 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and Bz). **EF-7412** acts an antagonist *in vivo* in pre- and postsynaptic 5-HT<sub>1A</sub> receptor sites and as an antagonist in dopamine D<sub>2</sub> receptor. © 1999 Elsevier Science Ltd. All rights reserved.

The discovery of multiple serotonin (5-HT) receptor subtypes in recent years has been accompanied by a parallel explosion in the development of drugs that alter 5-HT neurotransmission. The 5-HT<sub>1A</sub> receptor is the most intensively studied, as a result of it is involved in a variety of physiological and pathophysiological processes.<sup>1–3</sup> The 5-HT<sub>1A</sub> receptor belong to the class of G-protein coupled receptors (GPCRs) and the members of this class have a number of characteristic amino acid patterns in common, in spite of their different pharmacology. In particular, the transmembrane amino acid sequence of the 5-HT<sub>1A</sub> subtype is noteworthy for its high degree of homology to  $\alpha_1$ -adrenergic receptor subtypes,<sup>4</sup> so, a great number of ligands display affinity for both receptors. In this way, we have reported recently a new series of bicyclohydantoin- and imide-aryl piperazines **I,II**,<sup>5–9</sup> in which we have undertaken a systematic research of the structural factors that are responsible for 5-HT<sub>1A</sub>/ $\alpha_1$  selectivity. The study of the alkyl chain and the amide substructure have allowed us to suggest some important differences between the non-pharmacophoric sites of both receptors.<sup>9</sup> Regarding the aromatic ring substitution<sup>7</sup> it seems that *ortho*- and *meta*-positions are favorable for the affinity at both receptors, while *para*-substituted derivatives are practically inactive.

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In order to gain insight into the physicochemical influence of the 5-HT<sub>1A</sub>/α<sub>1</sub> receptor pharmacophores we have designed a training set of 32 compounds of general structure **III**. The amide moiety is a bicyclohydantoin or a diketopiperazine (X = -(CH<sub>2</sub>)<sub>3</sub>-, -(CH<sub>2</sub>)<sub>4</sub>-; m = 0, 1); the spacer length is 3 or 4 methylene units, which are the optimum values for both receptors, and the aromatic substituent R occupies the *ortho*- or *meta*- positions and it has been selected from a data base of 387 substituents using the EDISFAR program.<sup>10</sup> After the synthesis of the compounds and the evaluation of their affinities for 5-HT<sub>1A</sub> and α<sub>1</sub> receptors (most of the compounds showed high affinity for 5-HT<sub>1A</sub> and α<sub>1</sub> receptor binding site), we have carried out quantitative structure-activity relationships using artificial neural networks.<sup>11</sup>



The data set used was the *in vitro* 5-HT<sub>1A</sub> and α<sub>1</sub> receptor affinities (expressed as pK<sub>i</sub> values). Each compound was parametrized with six physicochemical descriptors (*F*, *R*, *V<sub>o</sub>*, *V<sub>m</sub>*, π<sub>*o*</sub>, π<sub>*m*</sub>) and three indicator variables (I<sub>A</sub> = 1 or 0 for X = -(CH<sub>2</sub>)<sub>4</sub>- or -(CH<sub>2</sub>)<sub>3</sub>-, I<sub>B</sub> = 1 or 0 for m = 1 or 0, I<sub>n</sub> = 1 or 0 for n = 4 or n = 3). The neural network employed for this modeling was a fully connected three layer network (input, hidden, output) trained by back-propagation of error. Initially the number of neurons in the input layer was equal to the number of molecular descriptors and the indicator variables, whereas the output layer had only one neuron. The number of neurons in the hidden layer was determined by trial and error. The best ANN models are shown in Table I. The dependence of biological activity on the physicochemical parameters was illustrated in 3-D diagrams. On the basis of the obtained plots, the 5-HT<sub>1A</sub> affinity has a nonlinear dependence with *F*, *V<sub>o</sub>*, *V<sub>m</sub>* and π<sub>*o*</sub>, nevertheless the nonlinear relationship is not far from the planar one. The α<sub>1</sub> affinity has a clear nonlinear dependence with *F*, *V<sub>o</sub>*, *V<sub>m</sub>*, π<sub>*o*</sub> and π<sub>*m*</sub>. A comparison of both analysis gives an additional understanding for 5-HT<sub>1A</sub>/α<sub>1</sub> selectivity: (a) High *F* values increase the binding affinity for 5-HT<sub>1A</sub> receptors and decrease the affinity for α<sub>1</sub> sites; (b) The lipophilicity at the *meta* position has only influence for the α<sub>1</sub> receptor; (c) The *meta* position seems to be implicated in the 5-HT<sub>1A</sub>/α<sub>1</sub> selectivity. While the 5-HT<sub>1A</sub> receptor is able to accommodate bulky substituents (about 60 Å<sup>3</sup>) in the region of

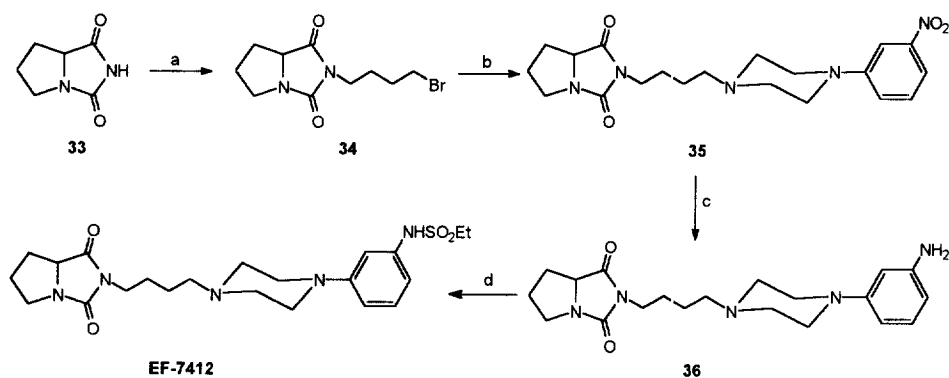
its active site, the steric requirements of the  $\alpha_1$  receptor at this position are more restricted (between 0–22 Å<sup>3</sup>). A good way to improve 5-HT<sub>1A</sub>/ $\alpha_1$  selectivity would be the synthesis of long chain derivatives bearing bulky substituents with high  $F$  values and low  $\pi$  values at the *meta* position. Among the different groups that fulfill these requirements the *m*-NHSO<sub>2</sub>Et was chosen ( $F = 0.419$ ,  $\pi_m = -0.64$ ,  $V_m = 65.31$ ). On this basis, the new ligand **EF-7412** ( $X = -(CH_2)_3-$ ,  $m = 0$ ,  $n = 4$ ,  $R = m\text{-NHSO}_2\text{Et}$ ) was designed and synthesized. This compound bound at 5-HT<sub>1A</sub> sites [ $K_{i\text{obsd}}$  (nM) =  $27 \pm 6$ ;  $K_{i\text{calcd}}$  (nM) = 36] and showed high selectivity over the  $\alpha_1$  receptor [ $K_{i\text{obsd}}$  (nM) >1000;  $K_{i\text{calcd}}$  (nM) = 2745].

**Table I.** ANN Models

Receptor	Non-significant Parameters	Architecture	r	r <sup>2</sup>	s
5-HT <sub>1A</sub>	$R$ , $\pi_m$	7-2-1	0.983	0.966	0.149
$\alpha_1$	$R$	8-2-1	0.991	0.982	0.136

Compound **EF-7412** was prepared as shown in Scheme 1. The reaction of the hydantoin **33**<sup>12</sup> with 1,4-dibromobutane in the presence of NaH in *N,N*-dimethylformamide (DMF) afforded the intermediate **34**.<sup>6</sup> Treatment of **34** with 1-(*m*-nitrophenyl)piperazine<sup>13</sup> yielded the nitroderivative **35**,<sup>7</sup> which was reduced to the amino compound **36**<sup>7</sup> by catalytic hydrogenation. The desired compound **EF-7412** was obtained by reaction of **36** with ethylsulfonyl chloride in the presence of pyridine in anhydrous acetone as solvent. Derivative **EF-7412** was characterized by IR and <sup>1</sup>H- and <sup>13</sup>C-NMR spectroscopy and gave satisfactory combustion analysis (C, H, N).<sup>14</sup> Hydrochloride salt of **EF-7412** was prepared as sample for biological assays.

On the other hand, binding affinities show that compound **EF-7412** possessed an appreciable affinity for D<sub>2</sub> receptor subtype ( $K_i = 22$  nM) and is selective for all other receptors examined (5-HT<sub>2A</sub>, 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and benzodiazepine Bz;  $K_i > 1000$  nM).



Reagents: (a) NaH, DMF, N<sub>2</sub>, 60 °C, 1 h, then Br(CH<sub>2</sub>)<sub>4</sub>Br, 110 °C, 2 h; (b) 1-(*m*-nitrophenyl)piperazine, Et<sub>3</sub>N, acetonitrile, 60 °C, 20 h; (c) H<sub>2</sub>/10% Pd(C), MeOH, 2 h; (d) EtSO<sub>2</sub>Cl, pyridine, acetone, N<sub>2</sub>, rt, overnight.

**Scheme 1**

Pharmacological evaluation of the activity of EF-7412 on 5-HT<sub>1A</sub> receptor function shows that subcutaneous administration of this compound did not alter rectal temperature neither the serotonergic syndrome (flat body posture and lower lip retraction) or plasma corticosterone levels. In addition, EF-7412 increased 5-HIAA/5HT and DOPAC/DA ratios in the mouse hypothalamus and slightly decreased spontaneous motor activity in the open field test. However, this compound blocked the hypothermia, the serotonergic syndrome, the enhancement of corticosterone secretion and the increase on 5-HT neuronal activity induced by 8-OH-DPAT. These results suggest that EF-7412 acts as an antagonist *in vivo* in pre- and postsynaptic 5-HT<sub>1A</sub> receptor sites. Furthermore, the fact that EF-7412 increased DA neuronal activity in the mouse hypothalamus and slightly decreased motor activity in rats (open field test) suggests that this compound may be acting as an antagonist in DA D<sub>2</sub> receptors.

To our knowledge, in this communication we describe the first selective derivative (EF-7412) with mixed 5-HT<sub>1A</sub>/D<sub>2</sub> antagonist properties and this derivative could be useful as a pharmacological tool.

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14. Data of EF-7412: 82% yield; mp 187–190 °C (methanol/ethyl ether); IR (CHCl<sub>3</sub>, cm<sup>-1</sup>) 3200, 1770, 1700; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.35 (t, *J* = 7.5 Hz, 3H, CH<sub>3</sub>), 1.50–1.72 (m, 5H, -(CH<sub>2</sub>)<sub>2</sub>-, H<sub>7</sub>), 2.05–2.13 (m, 2H, H<sub>6</sub>), 2.23–2.27 (m, 1H, H<sub>7</sub>), 2.41 (t, *J* = 7.5 Hz, 2H, CH<sub>2</sub>-Npip), 2.57 (t, *J* = 4.8 Hz, 4H, 2CH<sub>2</sub>-pip), 3.12 (q, *J* = 7.5 Hz, 2H, SO<sub>2</sub>CH<sub>2</sub>), 3.20 (t, *J* = 5.4 Hz, 4H, 2CH<sub>2</sub>-pip), 3.23–3.29 (m, 1H, H<sub>3</sub>), 3.50 (t, *J* = 7.2 Hz, 2H, NCH<sub>2</sub>), 3.69 (dt, *J* = 11.1, 7.2 Hz, 1H, H<sub>3</sub>), 4.09 (dd, *J* = 9.0, 7.5 Hz, 1H, H<sub>8a</sub>), 6.67–6.71 (m, 2H, H<sub>4</sub>- and H<sub>6</sub>-phenyl), 6.82 (t, *J* = 2.1 Hz, 1H, H<sub>2</sub>-phenyl), 7.18 (t, *J* = 8.4 Hz, 1H, H<sub>5</sub>-phenyl); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 8.2 (CH<sub>3</sub>), 23.6, 25.9 (-(CH<sub>2</sub>)<sub>2</sub>-), 26.9 (C<sub>6</sub>), 27.5 (C<sub>7</sub>), 38.6 (NCH<sub>2</sub>), 45.4 (C<sub>5</sub>, SO<sub>2</sub>CH<sub>2</sub>), 48.4 (2CH<sub>2</sub>-pip), 52.9 (2CH<sub>2</sub>-pip), 57.7 (CH<sub>2</sub>-Npip), 63.3 (C<sub>8a</sub>), 107.5 (C<sub>2</sub>-phenyl), 111.0, 112.1 (C<sub>4</sub>- and C<sub>6</sub>-phenyl), 130.0 (C<sub>3</sub>-phenyl), 137.9 (C<sub>3</sub>-phenyl), 152.2 (C<sub>1</sub>-phenyl), 160.8 (C<sub>3</sub>), 174.0 (C<sub>1</sub>). Anal. (C<sub>22</sub>H<sub>33</sub>N<sub>5</sub>O<sub>4</sub>S·2HCl·1/2H<sub>2</sub>O) C, H, N.